## **Research Note**

## Induction of polyembryony and secondary embryogenesis in culture for embryo rescue of stenospermocarpic genotypes of *Vitis vinifera* L.

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K e y w o r d s : polyembryony, seedless grapes, stenospermocarpy.

**Introduction:** In stenospermocarpic genotypes of *Vitis vinifera* L. it is possible to rescue the embryo and produce seedling plants by a procedure of *in vitro* embryo culture (CAIN *et al.* 1983; EMERSHAD and RAMMING 1984; SPIEGEL-ROY *et al.* 1985; GRAY *et al.* 1987; TSOLOVA 1990).

Multiple embryos resulting from this procedure and spontaneous polyembryony in culture have been occasionally reported (EMERSHAD and RAMMING 1984; DURHAM *et al.* 1989). Studies on spontaneous polyembryony in some seeded grapevine varieties and interspecific hybrids (*V. vinifera* x *V. riparia*) were undertaken by BOUQUET (1978, 1980, 1982). He has reported a low percentage (0.35 %) in *V. vinifera* and a higher rate (up to 7.7 %) polyembryony in interspecific hybrids.

Polyembryony is a classical means for naturally occurring haploids in angiosperms (PODUBNAIA-ARNOLDY 1976; BOUQUET 1980). Studies on the possibilities to induce polyembryony in culture of fertilized ovules of stenospermocarpic vine genotypes have not been made for ne time being.

The present research was undertakted with the following aims: 1. to study the possion has for induction of poltively high percentage of polyembryony in culture during *in vitro* embryo rescue of seedless grapevine varieties; 2. production of multiple embryos and maintaining in culture of secondary, tertiary, etc. embryogenesis.

**Materials and methods:** *Plant material:* Five selfpollinated newly selected seedless grapevine cultivars (Kishmish Moldavski, Kishmish Luchistii, Smirnoff, Seedless hybrid VI-4, Russalka 3) and 3 traditional cultivars (Sultanina, Kishmish Rozovii and Sultanina gigas) and 2 crosses (Seedless hybrid VI-4 x Corynth white and Smirnoff x Corynth white) growing in the ampelographic collection of the Institute of Viticulture and Enology, Pleven, Bulgaria were tested. Berries were collected 60 d after anthesis and stored in the dark at 4 °C in the presence of SO<sub>2</sub> for a period of 5 weeks. Sultanina gigas is propagated in Bulgaria under the name of Seedless white big. It has been identified only by ampelographic characteristics as Sultanina gigas (tetraploid form of Sultanina) by NEGRULY and KATEROV in the 1960ies (KATEROV and DONTCHEV, unpublished data). Embryo rescue and polyembryony induction: Ovules were aseptically separated and put in groups of 20 in petri dishes containing 20 ml of culture medium. The medium is NITSCH and NITSCH (1969) (NN69) supplemented with  $10^{-6}$  M Ga<sub>3</sub> (gibberellic acid),  $10^{-5}$  M IAA (indolyl acetic acid),  $10^{-6}$  M kinetin, 0.3 % active charcoal, 20 g/l saccharose and 0.8 g/l agar, adjusted to pH 5.7 before it was autoclaved. Different levels of vitamins and hormones in the induction medium were tested for Kishmish Moldavski, Russalka 3 and Sultanina gigas cultivars.

The ovules were cultured on induction medium for 12-14 weeks, then the undeveloped ovules were transferred on fresh induction medium. The germinating ovules were periodically counted, the first counting being executed after 8 weeks in culture, the last one after 23 weeks. The results presented in Tables 1 and 2 are the summarized data for each cultivar from the three countings after 8, 16 and 23 weeks in culture. The cultivation was performed at temperature of  $26\pm2$  °C and a photoperiod of 16/8 h.

Induction of secondary embryogenesis: The embryos, 10-15 mm in length, were transferred on NN69 growth regulator-free medium for development and induction of secondary embryogenesis in the zone between the hypocotyl and the root.

**Results and discussion:** The spontaneous polyembryony in grape might be due to several mechanisms (BOUQUET 1978, 1980, 1982; EM RSFAD and KAMMING 1984; DURHAM *et al.* 1989; EX ESTAD *et al.* 1989): 1) fertilizing and development of clore than one cell in the embryo sac; is ac tentive embryogenesis from the zygote and the poembryo; 3) embryogenic development of gametic cells in addition not be zygote in the embryo sac.

Possibilities for induction of polyembryony in the enospermocarpic grapevine genotypes used in our research were available due to the fact that there were proembryonal structures in the initial explant. According to BOUQUET (1980) the genetic determination and some external conditions like climate, physiological state of the grapevine plant, improved nourishment etc. were found to influence greatly the naturally occurring polyembryony.

In our experiment we achieved a high percentage (from 20.0 to 85.7, depending on the variety) of developing embryos producing polyembryonic clusters. The cluster percentage is lower from 1 to 11 if the datum is referred to the total number of the cultured ovules (Tab. 1; Figure, a).

The cultivars Kishmish Moldavski and Russalka 3, positively reacting to low temperature, were subjected to the influence of 2, 4, 6, 8 and 10 times increased concentration of vitamins and hormones and the cultivar Sultanina gigas to 3,5 and 9 times. It might be concluded that 4-5 times increasing of the vitamin and hormone level in the induction medium might increase the percentage of polyembryogenic clusters. The higher values suppressed the percentage of the rescued embryos in these variants.

In most of the studied genotypes we were able to induce secondary embryogenesis in the transition zone between the hypocotyl and the root (Figure, c) of embryos 12-15 mm in length (Tab. 1). The percentage of embryoids

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